Japan Academy Prize to:

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for "Regulatory Mechanism of Calcium Concentration inside the Cell"



Outline of the work:

When cells are stimulated IP₃ is produced (inside the cell) and subsequently the intracellular Ca²⁺ concentration increases - the later initiates/modulates a plethora of diverse physiological responses. It is now known that the IP₃ receptor regulates Ca²⁺ release from the endoplasmic reticulum, a major intracellular Ca²⁺ store, and hence plays an important role in living systems. However despite its importance, the mechanism through which IP₃ increases intracellular Ca²⁺ concentration was still unknown 20 years ago. Initially the 'IP₃ receptor' was hypothesized to be a target molecule of IP₃, and presumed to release Ca²⁺ from an unknown intracellular Ca²⁺ store. Dr. Mikoshiba's contribution to unraveling the mechanism of IP₃ mediated Ca²⁺ increase, was the identification of the IP₃ target molecule and its role as a Ca²⁺ release channel. His work on this subject originated from an analysis of the protein P400 (now named the IP₃ receptor), which is greatly decreased in the cerebellum of ataxic mutant mice. Dr. Mikoshiba started his studies with the purification of P400 and the generation of specific monoclonal antibodies against it. He then went on to make a key finding, by demonstrating that P400 is an IP₃ binding protein and a functional Ca²⁺ release channel he underlined P400 as the IP₃ receptor. Notably, the IP₃ receptor was shown to be located on the endoplasmic reticulum. Dr. Mikoshiba went on to determine the entire sequence of the IP₃ receptor, by cDNA cloning (1989).

Now that he had identified the IP₃ receptor, Dr Mikoshiba and colleagues focused their attention on obtaining an in-depth description of the relationship between IP₃ receptor structure and function. Analysis of the detailed structure of the IP₃ receptor identified it as an allosteric protein that changes its form (from windmill to square form) reversibly in the presence and absence of Ca^{2+} . These studies revealed a unique property of the IP₃ receptor, that even though its covalent bond is cleaved by trypsin into several peptides, they can reassemble to form functional IP₃-induced Ca^{2+} release channel. Dr. Mikoshiba's group also succeeded in conducting a three dimensional crystallographic analysis of both the IP₃ binding core and the regulatory region of the IP₃ receptor, and from this clarified the IP₃ receptor pore gating mechanism. The fresh knowledge was further utilized to construct a new IP₃ sensor, developed around state of the art FRET (fluorescence resonance energy transfer) technologies.

Dr. Mikoshiba's analysis, based on developmental biology, demonstrated that the IP₃ receptor is a Ca²⁺ oscillator and essential in: fertilization; dorso-ventral axis determination at the 4- to 8- cell stage after fertilization; cell division; and neurite extension. Furthermore IP₃ receptor type 1 deficient mice showed: cerebellar ataxia, demonstrating that the type 1 receptor is important for neuronal plasticity; an abnormal secretion of the nerve growth factor, BDNF. Comparatively, the analysis of IP₃ receptor type 2 and type 3 double deficient mice revealed an absence of exocrine function and a phenotype similar to that of Sjögren syndrome (which is characterized by dry eyes and dry mouth). Interestingly Sjögren syndrome patients were recently shown to have antibodies (in

their blood) against the IP₃ receptors.

Dr. Mikoshiba recently expanded the known role of the IP₃ receptor by the discovery of an endogenous pseudo IP₃, IRBIT (IP₃ receptor binding protein released with inositol 1,4,5-trisphosplate) that binds to the IP₃ binding core and is released following IP₃ application. IRBIT not only regulates the amplitude and frequency of Ca^{2+} oscillation, but also acts as a tertiary messenger - activating the Na⁺HCO⁻ cotransporter 1 and thereby regulating the acid-base balance. To re-iterate this important new finding, the function of IP₃ is not only to release Ca^{2+} but also to regulate acid-base balance. Furthermore, redox (oxydo-reduction) regulation was considered to be independent of Ca^{2+} signaling, but Dr. Mikoshiba linked the two signaling systems by discovering ERp44 as a new redox sensor that regulates the IP₃ receptor. In an effort to complete the list of IP₃ receptor interacting proteins, Dr. Mikoshiba has now screened the proteins associated with the IP₃ receptor and identified various molecules such as 4.1N, TRP channel, and glutamate receptors. These findings suggest that the IP₃ receptor is closely linked with many other signaling pathways and various diseases.

In conclusion, Dr. Mikoshiba and his collaborators, 20 years ago, discovered and sequenced the IP_3 receptor – an IP_3 gated Ca^{2+} channel that is located on the endoplasmic reticulum. He further revealed the detailed threedimensional structure by concerted studies involving X-ray crystallographic analysis and cryo-electron microscopy. Based on Dr. Mikoshiba's expertise in developmental biology and biochemistry and his unique and original ideas, he and his collaborators developed and advanced research on "Regulatory Mechanism of Calcium Concentration inside the Cell."

References

- Mikoshiba, K., Huchet, M., Changeux, J. P. (1979) Biochemical and immunological studies on the P400 protein, a protein characteristic of the Purkinje cell from mouse and rat cerebellum. Dev. Neurosci. 2(6): 254–275.
- Furuichi, T., Yoshikawa, S., Miyawaki, A., Wada, K., Maeda, N., Mikoshiba, K. (1989) Primary structure and functional expression of the inositol 1,4,5-trisphosphate-binding protein P400. Nature 342(6245): 32–38.
- Miyawaki, A., Furuichi, T., Maeda, N., Mikoshiba, K. (1990) Expressed cerebellar-type inositol 1,4,5-trisphosphate receptor, P400, has calcium release activity in a fibroblast L cell line. Neuron 5(1): 11–18.
- Maeda, N., Kawasaki, T., Nakade, S., Yokota, N., Taguchi, T., Kasai, M., Mikoshiba, K. (1991) Structural and functional characterization of inositol 1,4,5-trisphosphate receptor channel from mouse cerebellum. J. Biol. Chem. 266(2): 1109–1116.
- 5) Miyazaki, S., Yuzaki, M., Nakada, K., Shirakawa, H., Nakanishi, S., Nakade, S., Mikoshiba, K. (1992) Block of Ca²⁺ wave and Ca²⁺ oscillation by antibody to the inositol 1,4,5-trisphosphate receptor in fertilized hamster eggs. Science 257(5067): 251–255.
- Kume, S., Muto, A., Aruga, J., Nakagawa, T., Michikawa, T., Furuichi, T., Nakade, S., Okano, H., Mikoshiba, K. (1993) The *Xenopus* IP₃ receptor: structure, functiion, and localization in oocytes and eggs. Cell 73: 555–570.
- 7) Kume, S., Muto, A., Inoue, T., Suga, K., Okano, H., Mikoshiba, K. (1997) Role of inositol 1,4,5-trisphosphate receptor in ventral signaling in *Xenopus* embryos. Science 278(5345): 1940–1943.
- 8) Inoue, T., Kato, K., Kohda, K., Mikoshiba, K. (1998) Type 1 inositol 1,4,5-trisphosphate receptor is required for induction of long-term depression in cerebellar Purkinje neurons. J. Neurosci. 18(14): 5366–5373.
- 9) Takei, K., Shin, R. M., Inoue, T., Kato, K., Mikoshiba, K. (1998) Regulation of nerve growth mediated by inositol 1,4,5-trisphosphate receptors in growth cones. **Science** 282(5394): 1705–1708.
- 10) Michikawa, T., Hirota, J., Kawano, S., Hiraoka, M., Yamada, M., Furuichi, T., Mikoshiba, K. (1999)

Calmodulin mediates calcium-dependent inactivation of the cerebellar type 1 inositol1,4,5-trisphosphate receptor. **Neuron** 23: 799–808.

- Yoshikawa, F., Iwasaki, H., Michikawa, T., Furuichi, T., Mikoshiba, K. (1999) Trypsinized cerebellar inositol 1,4,5-trisphosphate receptor. Structural and functional coupling of cleaved ligand binding and channel domains. J. Biol. Chem. 274(1): 316–327.
- 12) Fujii, S., Matsumoto, M., Igarashi, K., Kato, H., Mikoshiba, K. (2000) Synaptic plasticity in hippocampal CA1 neurons of mice lacking type 1 inositol-1,4,5-trisphosphate receptors. Learn & Memory 7(5): 312–320.
- Nishiyama, M., Hong, K., Mikoshiba, K., Poo, M. M., Kato, K. (2000) Calcium stores regulate the polarity and input specificity of synaptic modification. Nature 408(6812): 584–588.
- 14) Ma, H. T., Patterson, R. L., van Rossum, D. B., Birnbaumer, L., Mikoshiba, K., Gill, D. L. (2000) Requirement of the inositol trisphosphate receptor for activation of store-operated Ca²⁺ channels. **Science** 287: 1647–1651.
- 15) Fukami, K., Nakao, K., Inoue, T., Kataoka, Y., Kurokawa, M., Fissore, R. A., Nakamura, K., Katsuki, M., Mikoshiba, K., Yoshida, N., Takenawa, T. (2001) Requirement of phospholipase Cδ4 for the zona pellucidainduced acrosome reaction. Science 292(5518): 920–923.
- 16) Bosanac, I., Alattia, J. R., Mal, T. K., Chan, J., Talarico, S., Tong, F. K., Tong, K. I., Yoshikawa, F., Furuichi, T., Iwai, M., Michikawa, T., Mikoshiba, K., Ikura, M. (2002) Structure of the inositol 1,4,5-trisphosphate receptor binding core in complex with its ligand. Nature 420(6916): 696–700.
- Saneyoshi, T., Kume, S., Amasaki, Y., Mikoshiba, K. (2002) The Wnt/calcium pathway activates NF-AT and promotes ventral cell fate in *Xenopus* embryos. Nature 417(6886): 295–299.
- 18) Nagai, T., Ibata, K., Park, E. S., Kubota, M., Mikoshiba, K., Miyawaki, A. (2002) A variant of yellow fluorescent protein with fast and efficient maturation for cell-biological applications. Nature Biotechnol. 20: 87–90.
- 19) Hamada, K., Terauchi, A., Mikoshiba, K. (2003) Three-dimensional rearrangements within inositol 1,4,5-trisphosphate receptor by calcium. J. Biol. Chem. 278(52): 52881–52889.
- 20) Bannai, H., Inoue, T., Nakayama, T., Hattori, M., Mikoshiba, K. (2004) Kinesin dependent, rapid, bi-directional transport of ER sub-compartment in dendrites of hippocampal neurons. J. Cell Sci. 117(Pt 2): 163–175.
- 21) Sato, C., Hamada, K., Ogura, T., Miyazawa, A., Iwasaki, K., Hiroaki, Y., Tani, K., Terauchi, A., Fujiyoshi, Y., Mikoshiba, K. (2004) Inositol 1,4,5-trisphosphate receptor contains multiple cavities and L-shaped ligandbinding domains. J. Mol. Biol. 336(1): 155–164.
- 22) Bosanac, I., Yamazaki, H., Matsu-Ura, T., Michikawa, T., Mikoshiba, K., Ikura, M. (2005) Crystal structure of the ligand binding suppressor domain of type 1 inositol 1,4,5-trisphosphate receptor. **Mol. Cell** 17(2): 193–203.
- 23) Futatsugi, A., Nakamura, T., Yamada, M. K., Ebisui, E., Nakamura, K., Uchida, K., Kitaguchi, T., Takahashi-Iwanaga, H., Noda, T., Aruga, J., Mikoshiba, K. (2005) IP₃ receptor types 2 and 3 mediate exocrine secretion underlying energy metabolism. Science 309(5744): 2232–2234.
- 24) Higo, T., Hattori, M., Nakamura, T., Natsume, T., Michikawa, T., Mikoshiba, K. (2005) Subtype-specific and ER lumenal environment-dependent regulation of inositol 1,4,5-trisphosphate receptor type 1 by ERp44. Cell 120(1): 85–98.
- 25) Ando, H., Mizutani, A., Kiefer, H., Tsuzurugi, D., Michikawa, T., Mikoshiba, K. (2006) IRBIT suppresses IP₃ receptor activity by competing with IP₃ for the common binding site on the IP₃ receptor. Mol. Cell 22(6): 795–806.
- 26) Shirakabe, K., Priori, G., Yamada, H., Ando, H., Horita, S., Fujita, T., Fujimoto, I., Mizutani, A., Seki, G., Mikoshiba, K. (2006) IRBIT, an inositol 1,4,5-trisphosphate receptor-binding protein, specifically binds to and activates pancreas-type Na⁺/HCO₃₋ cotransporter 1 (pNBC1). **Proc. Natl. Acad. Sci. USA** 103(25): 9542–9547.

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